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Somatic mutational landscapes of ductal and lobular breast carcinomas in the GENIE Cohort v8.1: real world actionability assessment in 8,756 patients

Alessandro Leal¹, Patricia Taranto¹, Poliana Bergamaschine Giovani Blasi¹, Bianca Baron Geronimo¹, Carlos Tadeu Garrote², Fernando Moura¹

¹Centro de Oncologia e Hematologia Einstein Dayan-Daycoval, Hospital Israelita Albert Einstein, Sao Paulo, Brazil; ²Instituto do Cancer do Estado de Sao Paulo, Sao Paulo, Brazil



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Background

Invasive ductal carcinoma (IDC) and invasive lobular carcinoma (ILC) of the breast typically present distinct clinicopathological characteristics and responsiveness to systemic therapy¹. In addition, breast cancer data from The Cancer Genome Atlas (TCGA) have shown these two pathological subtypes also present distinct genomic features when analyzed using DNA copy number arrays and whole exome sequencing platforms². More recently, the AACR Project GENIE Consortium, which is a publicly accessible international cancer registry of real-world data assembled through data sharing among leading cancer centers in the world, have allowed in-depth analyses of clinical actionability using patient-level data from clinical next-generation sequencing (NGS) assays³. In this study, we assessed the somatic mutational landscapes of a large cohort (n = 8,756) of invasive breast carcinomas from 19 institutions participating in the GENIE Consortium Cohort (v8.1) and examine clinical actionability of unique mutations identified in each breast cancer subtype.

Methods

We assessed the eighth data release of the GENIE Consortium Cohort v8.1 (Fig. 1) encompassing targeted sequencing data from 7,647 IDC and 1,109 ILC cases. Clinical features and somatic mutations including single-nucleotide variants, small indels, fusions, and copy number alterations (CNAs) were retrieved from cBioportal and SAGE Bionetworks. All patient samples were de-identified and encoded with GENIE sample codes. Gene actionability was examined using CIVIC, OncoKB, and ESCAT publicly available knowledgebases.

Results

Patients with IDC tumors were 5 years younger than patients with ILC tumors at the time sequencing data was reported (median 55 versus median 60, *Kruskal-Wallis*, $p < 10e-10$) (Fig. 2A). Both IDC and ILC had on average 2 mutations per tested sample. Overall, IDC and ILC tumors had median fractions of 22% and 14% of their genomes altered, respectively (*Kruskal-Wallis*, $p < 10e-10$) (Fig. 2B).

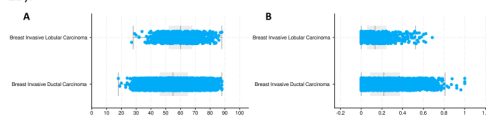


Fig. 2. Overview of reported breast cancer samples at GENIE Consortium Cohort v8.1. A) Age distribution at which sequencing was reported for invasive lobular carcinoma and invasive ductal carcinoma. B) Fraction of genome altered for invasive lobular carcinoma and invasive ductal carcinoma.

A gene enrichment analysis including 938 genes with point mutations and indels identified CDH1 (LR 4.66, $p < 1e-10$), RHOA (LR 2.81, $p = 1.3e-10$), PTK2B (LR 2.68, $p = 5.2e-4$), ERBB2 (LR 1.80, $p < 1e-10$), TBX3 (LR 1.72, $p < 1e-10$), FOXA1 (LR 1.49, $p = 2.5e-10$) and RUNX1 (LR 1.25, $p = 3.1e-9$) as genes significantly enriched in ILC tumors. On the other hand, mutations in GATA3 (LR = 1.67, $p < 1e-10$) and TP53 (LR = 1.55, $p < 1e-10$) were significantly enriched in IDC tumors (Fig. 3A, Table 1). A further gene enrichment analysis for copy-number alterations in 1139 genes showed amplification in PARP1 (LR 1.55 $p = 2.5e-3$) and deep deletions in IKZF1 (LR 2.8, $p = 2.2e-3$) and CDH1 (LR = 1.88, $p = 1.7e-4$) as the most enriched genes with CNAs in ILC. In parallel, amplifications in ERBB2 (LR 1.65, $p = 1e-10$), MYC (LR 1.64, $p = 1e-10$), COL22A1 (LR 1.19, $p = 1.6e-5$), BRIP1 (LR 2.66, $p < 1e-10$), CDK12 (LR 1.55, $p = 2.6e-9$), PPM1D (LR 3.1, $p < 1e-10$), RAD51C (LR 2.85, $p = 3.8e-8$), AURKA (LR 3.2, $p = 1e-8$) and deep deletion in CDKN2A (LR 2.1, $p = 1.9e-6$) were enriched in IDC tumors (Fig. 3B, Table 2).

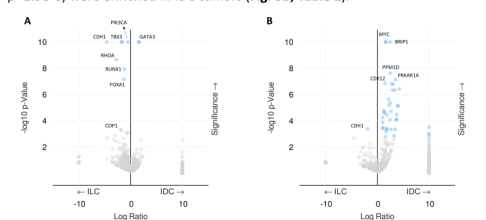


Fig. 3. Differential point mutations and copy number alterations (CNAs) at GENIE Consortium Cohort v8.1. A) Volcano plot showing the log ratio for point mutations across invasive lobular carcinoma (ILC) and invasive ductal carcinoma (IDC). B) Volcano plot showing the log ratios for CNAs across ILC and IDC.

Table 1. Genes harboring missense and nonsense mutations which are significantly enriched in invasive ductal carcinoma (blue) or invasive lobular carcinoma (red).

Gene	Cytoband	Invasive Ductal Carcinoma	Invasive Lobular Carcinoma	Log Ratio	p-Value	q-Value	Enriched In
CDH1	16q22.1	207 (3.06%)	631 (66.49%)	-4.44	<10e-10	<10e-10	Lobular
TP53	17p13.1	2907 (43.79%)	148 (15.37%)	1.49	<10e-10	<10e-10	Ductal
PIK3CA	3q26.32	2368 (34.90%)	466 (48.90%)	-0.52	<10e-10	<10e-10	Lobular
TBX3	12q24.31	197 (4.06%)	89 (22.90%)	-1.67	<10e-10	<10e-10	Lobular
ERBB2	17q12	199 (2.93%)	86 (9.02%)	-1.62	<10e-10	<10e-10	Lobular
GATA3	10p14	803 (13.47%)	44 (5.16%)	1.4	<10e-10	<10e-10	Ductal
RHOA	9q21.31	19 (0.31%)	22 (4.07%)	-2.74	5.86e-09	3.30e-06	Lobular
RUNX1	21q22.12	183 (3.07%)	62 (7.44%)	-1.22	3.81e-09	7.42e-06	Lobular
FOXA1	16q22.1	119 (1.79%)	47 (8.70%)	-1.2	2.40e-06	4.14e-04	Lobular
CFBR	16q22.1	214 (4.07%)	55 (7.30%)	-0.84	1.17e-04	0.0182	Lobular
BRIP1	17q23.2	104 (1.59%)	3 (0.36%)	2.33	3.41e-04	0.0489	Ductal

Table 2. Genes harboring amplifications or deep deletions which are significantly enriched in invasive ductal carcinoma (blue) or invasive lobular carcinoma (red).

Gene	Cytoband	Type	Invasive Ductal Carcinoma	Invasive Lobular Carcinoma	Log Ratio	p-Value	q-Value	Enriched In
ERBB2	17q12	Amp	826 (14.84%)	41 (4.88%)	1.56	<10e-10	<10e-10	Ductal
BRIP1	17q23.2	Amp	349 (6.01%)	9 (1.07%)	2.49	<10e-10	1.83e-09	Ductal
MYC	8q24.21	Amp	429 (10.90%)	23 (3.33%)	1.84	<10e-10	3.71e-08	Ductal
PPM1D	17q23.2	Amp	150 (6.49%)	4 (0.23%)	1.67	2.35e-08	6.57e-06	Ductal
PRKRA	17q24.2	Amp	149 (3.58%)	2 (0.31%)	1.54	7.09e-08	1.59e-05	Ductal
CDK12	17q12	Amp	316 (8.38%)	19 (3.36%)	1.48	1.41e-07	2.28e-05	Ductal
CDY98	17q23.3	Amp	162 (4.14%)	4 (0.61%)	2.75	1.32e-07	2.28e-05	Ductal
SPOP	17q23.33	Amp	147 (4.36%)	3 (0.53%)	3.04	1.43e-07	2.28e-05	Ductal
AURKA	17q24.1	Amp	108 (3.13%)	1 (0.19%)	1.24	3.18e-07	4.64e-05	Ductal
AURKA	17q24.1	Amp	143 (3.60%)	3 (0.46%)	1.97	4.53e-07	4.64e-05	Ductal
RMF43	17q22	Amp	126 (3.74%)	2 (0.35%)	3.4	4.37e-07	4.64e-05	Ductal
GNAS	20q13.12	Amp	165 (4.19%)	6 (0.91%)	2.2	2.22e-06	2.07e-04	Ductal
RAD51C	17q22	Amp	134 (4.04%)	4 (0.71%)	2.51	4.13e-06	3.57e-04	Ductal
GATA3	10p14	Amp	165 (2.37%)	5 (0.60%)	1.27	6.38e-06	4.95e-04	Ductal
ESFR	7q31.2	Amp	106 (1.84%)	1 (0.12%)	1.96	6.83e-06	4.95e-04	Ductal
CNE1	19q12	Amp	93 (2.89%)	1 (0.15%)	3.95	7.34e-06	5.14e-04	Ductal
COL22A1	8q24.23-q24.3	Amp	400 (21.89%)	18 (0.57%)	1.19	1.57e-05	1.04e-03	Ductal
NCOA2	20q13.12	Amp	131 (3.95%)	3 (0.49%)	2.64	1.90e-05	1.18e-03	Ductal
RAD21	8q24.11	Amp	252 (7.48%)	18 (3.13%)	1.56	2.30e-05	1.31e-03	Ductal
RECQL4	16q23.3	Amp	199 (5.59%)	12 (2.96%)	1.53	2.37e-05	1.31e-03	Ductal
IGFBP3	16q23.3	Amp	89 (2.12%)	1 (0.15%)	3.79	3.18e-05	1.31e-03	Ductal
CDKN2A	9p21.3	DeepDel	173 (8.00%)	1 (0.15%)	1.86	3.43e-05	1.85e-03	Ductal
IG	8q24.22	Amp	421 (22.62%)	21 (10.99%)	1.04	5.40e-05	2.73e-03	Ductal
PRDM1	6q21	Amp	77 (1.97%)	1 (0.15%)	3.68	7.70e-05	3.40e-03	Ductal
RTTL2	20q13.33	Amp	75 (4.21%)	1 (0.13%)	3.66	8.56e-05	3.40e-03	Ductal
GH1	17q23.3	Amp	158 (8.85%)	3 (1.60%)	2.44	8.06e-05	3.40e-03	Ductal
UBR5	6q23.3	Amp	392 (21.36%)	20 (10.54%)	1.02	1.38e-04	3.32e-03	Ductal
ROSL	6q23.1	Amp	59 (1.02%)	0 (0.00%)	>10	3.02e-04	0.0117	Ductal
STNAG	6q23.13	Amp	336 (18.06%)	16 (8.51%)	1.09	3.02e-04	0.0117	Ductal
AOC1	8q24.3	Amp	109 (6.12%)	5 (1.66%)	1.89	3.33e-04	0.0141	Ductal
DC44L2	8q23.3	Amp	35 (1.21%)	18 (5.57%)	1	3.97e-04	0.0141	Ductal
CDH1	16q22.1	DeepDel	24 (0.42%)	13 (1.42%)	-1.89	4.06e-04	0.0141	Lobular
CDKN2B	9p21.3	DeepDel	121 (3.88%)	6 (0.91%)	1.75	4.18e-04	0.0142	Ductal
IKZF3	17q13-q11.1	Amp	76 (1.24%)	2 (2.15%)	2.62	4.41e-04	0.0144	Ductal
SORL1	17q24.1	Amp	64 (1.17%)	1 (0.15%)	1.63	4.51e-04	0.0144	Ductal
PIK3CA	3q26.32	Amp	133 (2.31%)	4 (0.71%)	1.7	6.30e-04	0.0196	Ductal
NF1	17q11.2	Amp	51 (0.89%)	0 (0.00%)	>10	9.11e-04	0.0276	Ductal
ZNF127	12q12	Amp	47 (4.00%)	0 (0.00%)	>10	1.10e-04	0.0392	Ductal
PTPRD	9q21-q23	Amp	65 (1.24%)	1 (0.13%)	3.23	1.38e-03	0.0399	Ductal
BCAS3	17q23.2	Amp	169 (9.25%)	6 (1.19%)	1.54	1.42e-03	0.0454	Ductal

We identified 981 genes with point mutations across all 8,756 samples. From these, there are OncoKB curated information for 539 (54.9%) genes. Regarding variants and genes actionability for breast cancer, OncoKB and ESCAT present data for 16 (1.6%) and 11 (1.1%) genes, respectively (Table 3). Among enriched alterations for each histological subtype, the knowledgebase CIVIC does not present curated data available for genes TBX3, FOXA1, GATA3, COL22A1, BRIP1, PPM1D, and RAD51C. OncoKB only missed genes PTK2B and COL22A1.

Table 3. Genes harboring clinically actionable alterations in breast cancer at OncoKB and ESCAT.

Hugo Symbol	OncoKB Breast Cancer Data	OncoKB Breast Cancer Drug Use	OncoKB Breast Cancer Level	OncoKB Breast Cancer Number of Clusters	ESCAT Breast Cancer Data	ESCAT Tier
PIK3CA	Yes	Alpelisib + Fulvestrant	1	2	Yes	IA
AKT1	Yes	AZD5363	3A	4	Yes	IB
BRCA1	Yes	Talaporvicin, Olaparib	2	5	Yes	IA
BRCA2	Yes	Talaporvicin, Olaparib	2	4	Yes	IA
ERBB2	Yes	Lapatinib + Trastuzumab; Pertuzumab + Trastuzumab; Trastuzumab + Capecitabine; Trastuzumab + Ado-Trastuzumab; Emtricitabine; Capecitabine; Docetaxel	1	20	Yes	IA ou IB
ESR1	Yes	AZD5363 + Fulvestrant	3A	3	Yes	IA
NRX2	Yes	Larotrectinib + Entrectinib	1	7	Yes	IC
NRX3	Yes	Larotrectinib + Entrectinib	1	7	Yes	IC
PTEN	Yes	GSK25777 + AZD5363	4	2	Yes	IA
FGFR1	Yes	AZD4547; BGJ398; Erdafitinib; Dabrafenib	4	8	No	NA
FGFR2	Yes	AZD4547; BGJ398; Erdafitinib; Dabrafenib	4	8	No	NA
FGFR3	Yes	AZD4547; BGJ398; Erdafitinib; Dabrafenib	4	12	No	NA
KRAS	Yes	Cobimetinib; Binimetinib; Trametinib	4	8	No	NA
MET	Yes	Crizotinib	4	4	No	NA
MTOR	Yes	Temsirolimus; Everolimus	4	5	No	NA
NF1	Yes	Cobimetinib; Trametinib	4	5	No	NA
MDM2	No	NA	NA	NA	Yes	NA
NRX1	No	NA	NA	NA	Yes	IC

Conclusions

Real-world genomic data from the GENIE Consortium Cohort support that breast cancer presents distinct mutational landscapes for IDC and ILC tumors. For each histological subtype, we confirmed there are different levels of enrichments for shared mutations in actionable genes. Even though publicly available knowledgebases present curated information about commonly mutated genes in cancer, we noticed that actionability data for important cancer genes are still scarce.

References:

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Contact: alea@einstein.br